

October 2012

THE COLLEGE OF PHYSICIANS OF SOUTH AFRICA

R E G U L A T I O N S

**FOR ADMISSION TO THE EXAMINATION FOR THE
POST-SPECIALISATION**

SUB-SPECIALTY CERTIFICATE

IN

ADULT INFECTIOUS DISEASES

Cert ID(SA)Phys

1.0 BACKGROUND

- 1.1 Globally, infectious diseases constitute the single largest burden of illness afflicting mankind, especially in poorer regions of the world. The impact and future of the HIV and tuberculosis pandemics, and of novel, emerging and re-emerging infectious diseases necessitates the training of infectious diseases (ID) experts for the continent
- 1.2 This document includes the training programmes designed to produce competent ID sub-specialists with expertise in clinical infectious diseases, microbiology/virology and infection prevention and control (IPC)
- 1.3 Their training will combine clinical experience in infectious diseases medicine with laboratory training in microbiology and virology, and will provide exposure to principles of communicable diseases epidemiology, infection prevention and control and tropical public health. The ID sub-specialists will have true expertise in all aspects of diagnosis and management of organ, organ system and organism-specific infections
- 1.4 Infectious Diseases is an integrative clinical sub-speciality that draws upon not only all of the elements of general internal medicine and paediatrics, but also has relevance to all fields of medicine and surgery. It is integrally involved with the microbiology and epidemiology of infectious diseases. The ID specialist is a uniquely trained clinician fully capable of providing effective, ambulatory-based medicine and hospital-based clinical consultation and management. Furthermore, with thorough knowledge in infectious diseases epidemiology these ID specialists will be able to interact effectively with public health colleagues in such critical areas as HIV primary care education and training for general practitioners, and communicable disease outbreak investigation and control. Finally, trained ID specialists will also be able to perform invaluable roles in both the public and private sectors in hospital epidemiology/infection prevention and control, as well as rational antimicrobial utilisation

2.0 ELIGIBILITY

2.1 Entry is via Internal Medicine. Entrants must have passed FCP(SA).

2.2 Grandfather Clause:

Registration as an ID specialist under the Grandfather clause ended on 31st December 2010, without exception

3.0 TRAINING REQUIREMENTS**3.1 Scope and duration:**

Two years training is required comprising 6 months in the laboratory and 18 months in clinical ID work. The laboratory time need not be continuous

3.2 Overview of training requirements:

Clinical ID training must include in-patient and ambulatory care. Patient care must include both consultation and “hands-on” management. “Hands-on” management includes history-taking, physical examination, appropriate imaging and laboratory tests and appropriate bed-side investigations. The specialist will be able to prescribe and monitor antimicrobial therapy and should have sufficient insight into other forms of medical therapy such as immunosuppression. Practical experience in hospital infection prevention and control must form an integral part of ID training. For physicians/paediatricians, laboratory training includes clinical microbiology and virology. Research and scholarly activities will be emphasised

3.2.1 Training goals include the following:-

- The primary goal of the ID training is to educate and train competent, caring and compassionate ID sub-specialists who will provide quality care to patients
- ID sub-specialists must have a thorough knowledge of clinical microbiology and virology
- Ability to play a leadership role in hospital infection prevention and control, and antimicrobial policy formulation
- Insight into the principles and practice of communicable diseases epidemiology and control
- Trainees will be required to develop teaching and communication skills relevant to all levels of health care
- Trainees will be required to exhibit competence in research
- Critical appraisal of relevant scientific literature

3.3.../

3.3 Programme Content:

3.3.1 Although laboratory and clinical components of the 2-year programme are presented separately below, it is intended that whether laboratory time runs as a continuous block or not, that the training be integrated throughout the 2 year period so that there is constant interplay between clinical and laboratory elements of the Specialist's training.

3.3.1.1 Laboratory Microbiology, Virology and Infection Prevention Control (6 months / 25% total training time):

Provided by approved diagnostic microbiology/virology laboratories, with the goal of providing core and specialised knowledge in medical microbiology, virology and infection prevention and control. Trainees will become adept at critically interpreting laboratory data

3.3.1.1.1 Requirements:

- Microbiology, including infection prevention control and immunology of up to 5 months (minimum 4 months) and virology of up to 2 months (minimum 1 months)
- During this rotation, the trainee will also be exposed to mycology, parasitology, epidemiology, hospital-based infection prevention and control

3.3.1.2 Clinical Infectious Diseases (18 months):

The programme will provide training in adult infectious diseases in both in-patient and ambulatory settings. Clinical ID training will include consulting service at the accredited hospital(s) as well as "hands-on" management covering a wide variety of infectious diseases, under the guidance of ID faculty. Ambulatory care of patients with HIV will be emphasised by active participation in the longitudinal care of these individuals in HIV clinics.

3.3.1.3 Research and other academic activities:

3.3.1.3.1 Participation in one or more research projects in infectious diseases should be encouraged

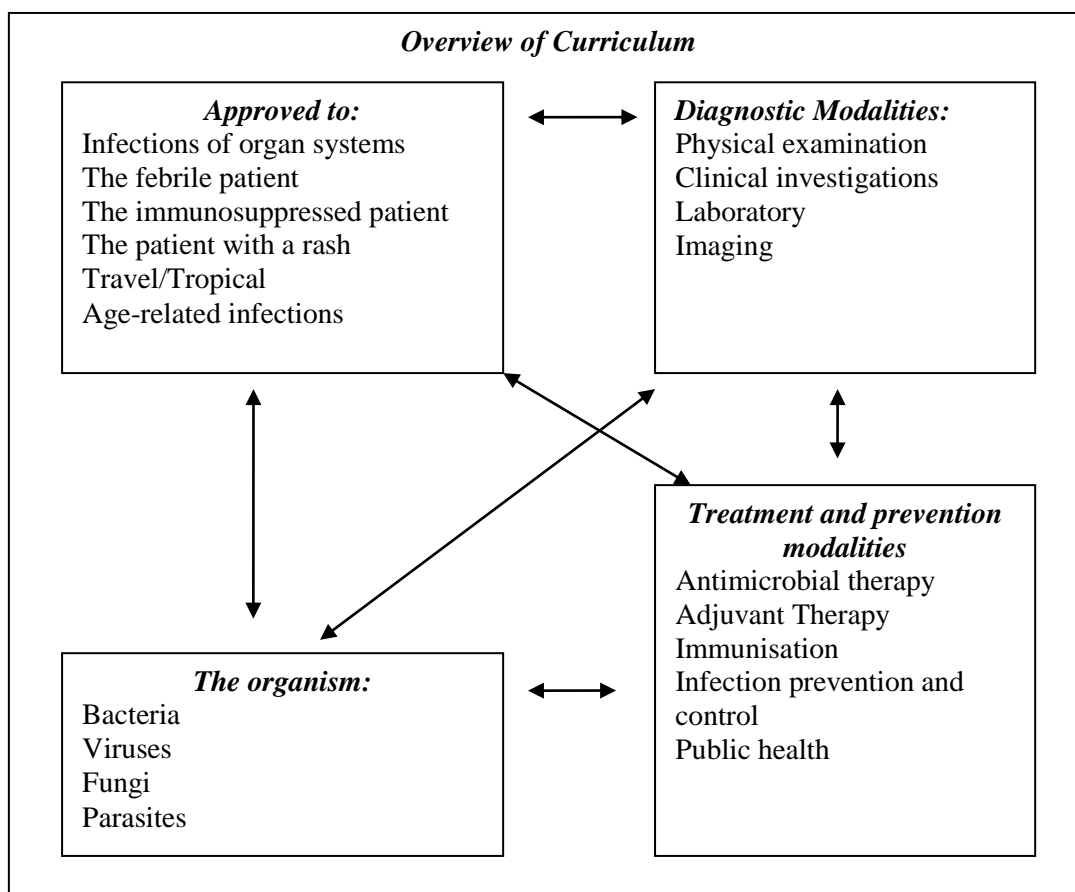
3.3.1.3.2 Publication of case reports, case series and research projects will be encouraged

3.3.1.3.3 Competence in the critical appraisal of scientific literature will be evaluated through journal club presentations, for example:

- Trainees will be required to participate in departmental meetings, presentations and teaching, and should be encouraged to attend conferences

3.4.../

3.4 Overview of Curriculum:



Guidelines are given in the Appendices 1 and 2. These are non-inclusive recommendations

4.0 EVALUATION

4.1 Portfolio:

4.1.1 Each candidate should maintain a portfolio (Appendix C), documenting the following:

- Details of cases managed
- Case presentations
 - by the candidate
 - case presentations attended
- Laboratory experience
 - hours spent in each section
 - techniques and experience gained
- “Elective” experience eg Immunology

4.1.2 Continuing in-course evaluation and feedback, (ie mentorship) is mandatory. Review of portfolio every 6 months by Head of the training Department/Unit is required. The portfolio must be signed off by trainee and trainer at each 6 month review.

4.2 Exit examination:

Prior to an exit evaluation of professional competence, each candidate's portfolio needs approval by the Head of the Infectious Diseases Unit. A letter of approval should be submitted to the CMSA

4.2.1 Final evaluation of professional competence must include:

- 3-hour written examination – Short answer essay paper (6 x 30 minute question)

Candidates who successfully pass the written examination will be invited to attend the next stage of the examination, which includes the OSCE and Clinical exam.

- 3-hour OSCE – this will include case histories, clinical and radiological scenarios, laboratory specimens
- Clinical examination – Consisting of 4 paper-based short cases (from the August/October 2010 examinations) – Each candidate will be examined by 2 sets of examiners, each set examining 2 cases. Each pair of examiners will contain an ID specialist and a laboratory specialist as a minimum.
- Review of the case portfolio will also form part of the evaluation
- Assessment of ability to perform research or at least to scrutinise and appropriately evaluate research data and scientific articles will form part of the
- This examination will be held under the auspices of CMSA twice a year

4.2.2 A sub-minimum mark of 50% is applied to each of the 3 components of the examination. The candidate must therefore gain 50% or more on each component to pass the examination. Any candidate scoring <50% for any of the 3 components will fail the examination. There is no sub-minimum applied to individual questions in any of the 3 components of the examination.

APPENDIX A

1.0 CLINICAL ID AND INFECTION PREVENTION CONTROL TRAINING FOR INTERNAL MEDICINE:

1.1 This list is not exhaustive or complete

- 1.2
- The following aspects should receive attention:
 - Epidemiology
 - Clinical presentation
 - Pathophysiology
 - Microbiology
 - Virology
 - Differential diagnosis
 - Investigations
 - Management
 - Drug interactions
 - Sepsis syndrome
 - Other treatment modalities in infections:
 - probiotics
 - gamma-globulins
 - exchange transfusion
 - Diseases of organ systems
 - Immunocompromised patients:
 - primary
 - secondary
 - HIV
 - Tuberculosis and other mycobacteria
 - Fungal infections
 - Sexually transmitted diseases
 - Hospital-acquired and health care-related infections
 - Diseases often mimicking and/or predisposing to infections:
 - auto-immune diseases
 - malignancies
 - Immunisations
 - Zoonoses
 - Geohelminth and other parasitic infestations
 - Travel-related infections
 - Tropical infections
 - Scenarios:
 - fever of unknown origin
 - recurrent infection
 - eosinophilia
 - antimicrobial and antiviral resistance
 - exanthemas – measles, chickenpox etc
 - Infections related to trauma including burns and human or animal bites
 - Antimicrobials:
 - drugs
 - pharmacology
 - toxicity
 - Infections in Geriatric patients and parenteral drug users
 - Para-infectious diseases including GBS, Chronic fatigue syndrome/ME, TP

1.3 Procedures:

- 1.3.1 Understand the indications for, be able to perform, and be able to interpret the results of the following clinical procedures:
- Thick and thin films for malaria
 - Filarial skin snips
 - Schistosoma rectal snips
 - Leprosy skin slit smears

2.0 INFECTION PREVENTION CONTROL TRAINING

2.1 Understand the principles and management of the following:

- Standard transmission
- Surveillance
- Hospital-Acquired Infections (HAI)
- HAIs in immunocompromised
- Practices and controversies in HAI
- Device-related HAI
- HAI audits
- HAI pathogen typing techniques
- Surgical site infections
- Outbreak response
- Tuberculosis IPC
- Sterilization and disinfection
- Antibiotic stewardship
- Structure and function of IPC advisory committees

2.2 ID specialist trainees must:

- Have had first hand experience of local infection control problems, including, outbreaks of infection and their management
- Be familiar with the workings of infection control meetings including local and regional infection control committees
- Be aware of those areas of hospital and community health that require infection control policies
- Have worked closely with the infection control nurse both in day-to-day duties and in the education of those involved with infection control issues
- Have participated in visits to clinical and non-clinical areas to advise on infection control. These should include kitchen inspections especially those conducted by environmental health officers. Relationship should be developed with key personnel in the CSSD, pharmacy and laundry
- Have insight into public health microbiology
- Have had some experience of communicable disease control in the community working with a CDC and Environmental Health Officers

APPENDIX B

1. LABORATORY TRAINING:

1.1 General Outline:

1.1.1 Objectives:

During the 6 months training, the ID subspeciality trainee, should develop:

- Specialised factual knowledge of the natural history of those diseases upon which the discipline of clinical microbiology and infectious diseases is based
- Interpretative skills so that a clinically useful opinion can be derived from laboratory data
- Technical knowledge gained from close acquaintance with laboratory technology, so that methodology appropriate to a clinical problem can be chosen, and so that quality control and quality assurance procedures can be implemented
- Research and development experience:
 - original thought and critical assessment of published work are important to allow the trainee to contribute in a team, and individually, to development of the service
- The long-life habits of reading, literature-searches, consultation with colleagues, attendance at scientific meetings, and the presentation of scientific work as part of continuing medical education (CME)
- Data management skills to evaluate information derived from the population served and from the technical procedures applied in the laboratory. These skills include familiarity with IT and the use of spreadsheets, databases and statistical packages, etc
- Management and communication skills:
 - the trainee must gain experience, under supervision, in planning departmental policies and develop the leadership skills necessary to implement them
- Familiarity with health and safety requirements for laboratories

1.2 Core curriculum:

This document sets out a curriculum, which covers the scientific base of clinical microbiology, as well as applied aspects of the subjects, including related fields such as infectious diseases, hospital epidemiology, and communicable diseases control

1.3 Training programme:

1.3.1 ID subspeciality trainees should have an understanding of the principles of the following, together with how they may be applied and research problems:

- Microbial structure, physiology and genetics
- Microbial taxonomy, classification and typing methods
- Host defence mechanisms, the immune system and immunity to infection
- Microbial pathogenicity
- Epidemiology of infectious diseases – their surveillance and control
- Antimicrobial agents, their mode of action and mechanisms of microbial resistance

1.4 Laboratory safety:

1.4.1 Prior to any 'hands on' experience of laboratory work, the ID subspeciality trainee should be instructed in basic safety requirements including correct laboratory dress and laboratory hygiene. Instruction should also be given on the immediate handling and disposal of specimens and contaminated articles (eg inoculating loops, pipettes) at the

laboratory bench, the dangers of aerosols and the procedure for dealing with spillages. At the end of formal training, the ID subspeciality trainee should be knowledgeable of:

- Local procedures for.../

- Local procedures for the safe transport of specimens or cultures and also with national and international postal and packaging regulations for such material
- Current knowledge of the regulations for hazardous biological agents
- The principles and operation of microbiological safety cabinets and the procedures for their decontamination and monitoring of air flow
- Infection control in the laboratory

1.5 Sterilisation and disinfection:

The trainee should understand the principles and uses of sterilisation and disinfection procedures and familiarity with microbiological waste disposal. ID subspeciality trainees should be familiar with methods of monitoring and be capable of guidance and disinfection in the laboratory, hospital or community

1.6 Handling of specimens:

1.6.1 At the end of formal training, the ID subspeciality trainee should:

- Be aware, for each specimen type, of the optimal methods for collection, transport (including transport media), storage, reception, identification and documentation, including the requirements for high-risk specimens. He/she should develop a sense of the continuity of identification of specimens from collection, through culture and further testing to the issuing of a final report. He/she needs to be aware of critical points in processing where the continuity may fail and be able to minimise the risk of this
- Be able to assess degrees of urgency for the processing of specimens, including the provision for an out of hours service and the communication of preliminary results as applicable
- Be able to assess when further testing or processing of a specimen is appropriate
- Be aware of existing reference facilities and their appropriate use

1.7 Microscopy:

- Understand the principles of light, phase-contrast, fluorescent and electron microscopy and be able to set up a light microscope
- Be able to perform routine staining techniques including fluorescent dyes
- Be familiar with the appearance of stained preparations and be able to recognise artifacts and their possible origin

1.8 Culture methods:

- Have a basic understanding of the diversity of microbial metabolism
- Be aware of the wide range of selective, enrichments and inhibitory media available for general and specialised use
- Be familiar with physical growth requirements of micro-organisms including atmosphere and optimal temperature and have an appreciation of the growth kinetics of both solid phase and broth cultures. Know those micro-organisms and clinical situations in which detectable growth may require prolonged incubations

- Understand the principles enabling one to process all common specimens, recognise potential pathogens from a mixture of colonies on culture plates, separate such colonies in order to achieve the growth necessary for further work

1.9 Further processing of cultures:

- Be aware of tests leading to the identification of all common pathogens including commercially produced kits (eg kits for enzyme assays) and rapid diagnostic kits, ELISA, latex agglutination
- Be aware of available reference facilities for further identification including serotyping and all other typing schemes both phenotypic and genotypic

1.10.../

1.10 Antimicrobial investigations:

- Be able to interpret the antibiotic sensitivities of an isolate using the common techniques
- Be able to interpret MIC and MBC tests as appropriate
- Have an understanding of antimicrobial assays and their relationship to the therapeutic and toxic effects on a patient and be able to advise on dosage regimens accordingly

1.11 Molecular technologies:

- Be aware of all major molecular technologies available in nucleic acid-based (molecular) techniques (eg RFLPs, hybridisation, macrorestriction analysis, LAMP, LCR, PCR and variations based on PCR)

1.12 Data handling:

- Have a basic understanding of information technology and in particular, computerised data handling. He/she should have an appreciation of the advantages and disadvantages of such systems and a basic understanding of the need for data protection and the Data Protection Act; and
- Be aware of available technologies for data broadcasting (eg EPINET)

1.13 Virology:

- Basic diagnostic virology methodology
- Interpretation of results, for both clinical and infection control purposes
- Virology policies in relation to health care workers, pregnancy, transplantation and immunisation
- When to refer to or request specialist virological expertise

1.14 Immunology:

- Work-up of suspected immunodeficiencies
- Interpretation of laboratory tests for major immunodeficiencies

PORTFOLIO OF LEARNING

CONTENTS

- SECTION 1** Certification of 6 Monthly Portfolio Review
- SECTION 2** Syllabus for the Cert ID(SA) Paed
- SECTION 3** Summary of Training
- SECTION 4** Candidate Details
- SECTION 5** Laboratory Training
- SECTION 6** Certification of Laboratory Training
- SECTION 7** Details of Cases Managed
- SECTION 8** Post-Graduate Lectures, Meetings, Workshops, Seminars, Symposia and Congresses
- SECTION 9** Reading and Research
- SECTION 10** Declaration on Completion of Training

Additional pages and supporting documentation should be attached, as necessary.

Please read the Regulations and Curriculum for the Cert ID(SA)Phys examination at the start of training – details are available on the CMSA website or a hard copy can be obtained from the CMSA office or your Programme Supervisor.

SECTION 1

CERTIFICATION OF 6 MONTHLY PORTFOLIO REVIEW

Date of Formative Assessments/Portfolio Review	Signature of Supervisor	Signature of Candidate	Comment

ELECTRONIC LINK TO COLLEGE REGULATIONS

[http://www.collegemedsa.ac.za/force_download.aspx?documentid=191&Name=Cert ID\(SA\) Phys Regulations](http://www.collegemedsa.ac.za/force_download.aspx?documentid=191&Name=Cert ID(SA) Phys Regulations)

SECTION 3

CANDIDATE DETAILS

SURNAME:.....

FIRST NAMES:.....

ID NUMBER:.....

HPCSA NUMBER:.....

TRAINEE POST NUMBER:

WORK ADDRESS:

.....

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RESIDENTIAL ADDRESS:

.....

.....

PREFERRED POSTAL ADDRESS:

.....

.....

EMAIL ADDRESS:.....

TELEPHONE NUMBER: (Work):.....(Home):.....

CELLPHONE NUMBER:.....

FAX NUMBER:

UNDERGRADUATE MEDICAL QUALIFICATIONS

UNIVERSITY: YEAR:.....

INTERNSHIP

HOSPITAL: YEAR:.....

COMMUNITY SERVICE

HOSPITAL: YEAR:.....

EMPLOYMENT HISTORY

INSTITUTION: YEAR:.....

INSTITUTION: YEAR:.....

INSTITUTION: YEAR:.....

OTHER REGISTERABLE POST-GRADUATE QUALIFICATIONS

DIPLOMA/DEGREE: YEAR:.....

INSTITUTION:

DIPLOMA/DEGREE: YEAR:.....

INSTITUTION:

DIPLOMA/DEGREE: YEAR:.....

INSTITUTION:

RELEVANT DETAILS / EXPERIENCE RELATING TO ADULT INFECTIOUS DISEASES

Prior to commencement of Cert ID(SA) Phys training

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SECTION 5

LABORATORY TRAINING

Record of Laboratory Procedures

For all the processes / procedures listed, the most important component is understanding the principles of the test, and the limitations of the test. Candidates are NOT expected to be competent to perform the tests in a laboratory setting. While performing the tests in a training environment will most probably facilitate understanding of the principles, the listed recommendations of how many tests to perform and/or observe is meant to be a guideline. Refer to appendix B of the Regulations for admission to the Examination for the sub-specialty Certificate in Infectious Diseases of the College of Physicians of South Africa.

1.0 Bacterial identification and susceptibility testing:

Procedure	Requirement	Trained by	Date
Perform and read Gram stain	Perform minimum 10 Including complicated plates		
Reading plates (macroscopic description and identification of bacterial colonies)	Perform minimum 10		
Bacterial identification: Catalase, DNase, oxidase, sugars	Understand principles,		
Bacterial identification: automated methods	Understand principles, observe at least 5		
Antimicrobial susceptibility testing methods: Prepare and interpret E test	Perform minimum 5		
Disc diffusion	Perform minimum 5		
Cut off plate	Understand principles		
Broth dilution	Understand principles		
Hodge test (carbapenemase production)	Understand principles		
ESBL identification	Understand principles		
Automated sensitivity	Understand principles		

2.0 Specimen processing

Procedure	Requirement	Trained by	Date
Sterile specimen collection	Understand principles		
Automated blood culture systems	Understand principles		
Process positive blood culture bottle	Perform minimum 5		
Process sputum specimen (smear, Bartlett score, inoculate plates)	Perform minimum 5		
Process CSF specimen (gram, cell count, inoculate plates)	Perform minimum 5		
Process stool specimen (Wet prep, iodine stain, auramine stain, inoculate plates)	Perform minimum 5		
Process urine specimen (microscopy cell count, inoculate plates)	Perform minimum 5		
Process pus swabs (gram, inoculate plates)	Perform minimum 5		
Process fungal cultures (inoculate plates; macroscopic description of colonies; microscopy)	Perform minimum 5		

3.0 TB specimens

Procedure	Requirement	Trained by	Date
Ziehl Neelsen stain and microscopy	Perform minimum 5		
Auramine stain and microscopy	Perform minimum 5		
Process specimen for culture (including decontamination)	Understand principles, observe at least 5		
Process specimen for molecular testing (GeneXpert, line probe assay)	Understand principles		
Interpret results of TB molecular tests	Understand principles, observe at least 10		
TB phenotypic sensitivity testing	Understand principles, observe at least 5		
Identification of non-tuberculous mycobacteria (phenotypic appearance and PCR testing)	Understand principles		

4.0 Molecular Biology Methods

Procedure	Requirement	Trained by	Date
Nucleic Acid Extraction Manual (manual and automated)	Understand principles, and troubleshooting		
Preparation of master mixes	Understand principles, and troubleshooting		
Use of thermal cyclers	Understand principles		
Gel Electrophoresis	Understand principles, and troubleshooting		
Real-Time PCR	Understand principles, and troubleshooting		
Sequence analysis	Understand principles		
HIV drug resistance testing	Understand principles		
Molecular typing of organisms	Understand principles		
Multiplex PCR	Understand principles		

5.0 Serology

Procedure	Requirement	Trained by	Date
Syphilis serology – perform and interpret RPR, immunofluorescent assays	Perform minimum 5 and troubleshooting		
Automated ELISA - indications, interpretation of results	Understand principles, observe at least 5		
Manual ELISA – indications, interpretation of results	Understand principles, observe at least 5		
IgG avidity assays	Understand principles		
Rapid tests - immuno-chromatographic	Understand principles		
Rapid tests - particle agglutination	Understand principles		

6.0 Virology

Procedure	Requirement	Trained by	Date
Automated HIV PCR testing	Understand principles, and troubleshooting		
Automated HIV viral load testing	Understand principles, and troubleshooting		
EBV / CMV viral load	Understand principles		
CMV pp65 antigenaemia	Understand principles		
Virus culture, identification by cytopathic effect	Understand principles		
Virus identification/typing by neutralisation	Understand principles		
Antibody detection by neutralisation	Understand principles		
Virus storage/ retrieval	Understand principles		
Influenza typing by HAI	Understand principles		

7.0 Immunology laboratory

Procedure	Requirement	Trained by	Date
Lymphocyte subset analysis	Understand principles, and interpretation		
Total immunoglobulins and subclasses	Understand principles		
Neutrophil burst test	Understand principles		
Serum protein electrophoresis	Understand principles		
Total complement, individual component assay	Understand principles		

8.0 Laboratory safety and management

Skill	Trained by	Date
Biosafety levels, safe handling of samples from a patient with suspected or proven viral haemorrhagic fever		
Decontamination of environment following a spill		
Safe handling of sharps, human material, hazardous waste		
Quality assurance and laboratory accreditation		

9.0 Infection Control and Prevention

Skill	Trained by	Date
Investigation of an outbreak / unusual cluster of cases		
Transmission based precautions, hand hygiene		
Principles of disinfection and sterilization		
Visit sterilization unit		
Visit hospital kitchen / milk kitchen		
Antibiotic stewardship / analysis of bacterial susceptibility surveillance data		

SECTION 6

CERTIFICATION OF LABORATORY TRAINING

1.0 Microbiology (Including Immunology)

I,(Head: Department of Microbiology)
hereby declare that the candidate
has satisfactorily completed his/her laboratory training, from
to as required for the CertID(SA) Phys.

Date:

.....

Signature

2.0 VIROLOGY

I,(Head: Department of Virology) hereby
declare that the candidate has
satisfactorily completed his/her laboratory training, from
to as required for the CertID(SA) Phys.

Date:

.....

Signature

3.0 INFECTION PREVENTION CONTROL (IPC)

I,(Head: Department of IPC) hereby declare
that the candidate has
satisfactorily completed his/her
IPC training, from to as
required for the CertID(SA) Phys.

Date:

.....

Signature

SECTION 7

DETAIL OF CASES MANAGED

Provide information on a maximum of **ten** cases per disease-topic. The purpose is to demonstrate that complicated cases were managed and that an adequate spectrum of diseases was encountered. Refer to appendix A of the Regulations for admission to the Examination for the sub-specialty Certificate in Infectious Diseases of the College of Physicians of South Africa.

Examples of conditions

Fever of Unknown Origin

Fever in immunocompromised host

Hospital acquired Infection in the ICU

Complicated Malaria

Rickettsial Infections

Leptospirosis

Surgical infections

Infective endocarditis

Osteomyelitis

Septic arthritis

Drug resistant tuberculosis

Typhoid fever

Pneumocystis pneumonia

Cryptococcosis

Herpes virus infections

Cytomegalovirus infections

Complicated

parvovirus infections

Cryptosporidiosis

Influenza infections

Hepatitis B

Rabies

HIV drug resistance

Post exposure prophylaxis

Travel advice



SECTION 10

DECLARATION ON COMPLETION OF TRAINING

I,hereby do solemnly declare that all information contained in this PORTFOLIO OF LEARNING is a true and accurate record of my professional experience, education and training from to representing the period of training for the Cert ID(SA) Paed.

Signature of Candidate:

Name of Candidate:Date:

Trainee Number:

Signature: Head of Training Unit:

Name:Date:

Signature: Head of Academic Department:.....

Name:Date: